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J. Fuentes^a; T. Cuevas^a; M. A. Pradera^a

^a Departamento de Química Orgánica, Facultad de Química, Universidad de Sevilla, Sevilla, Spain

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REGIOSELECTIVE BENZOYLATION OF N-PROTECTED D-GLUCOSAMINE

J. Fuentes*, T. Cuevas and M. A. Pradera

Departamento de Química Orgánica, Facultad de Química,
Universidad de Sevilla, Apartado 553, 41071 Sevilla, Spain

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ABSTRACT

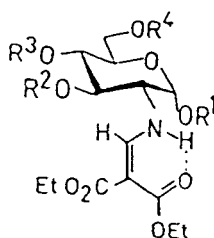
Regioselective benzoylations of 2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (1) yielded 6-mono-O- (2) 3,6-di-O- (3), 4,6-di-O- (4) and 1,3,6-tri-O-benzoyl- (5) derivatives. The fully benzoylated compound 6 was the major product when excess of benzoyl chloride was used. The di-O-acetyl derivatives (7 and 8) of 3 and 4 respectively were also prepared. The structures of 2-8 are based on analytical and/or spectroscopic data. Some data on MS fragmentation pathways of the partially protected compounds 2 and 4 are also reported.

INTRODUCTION

Selectively esterified monosaccharide derivatives are useful precursors for specific functionalisation¹⁻² and for oligosaccharide syntheses.³ On the other hand, D-glucosamine is a frequent structural unit in natural oligo- and polysaccharides. Regioselective acylation of sugar derivatives has been performed by both chemical^{1,2,4} and enzymatic⁵⁻⁸ methods and regioselective benzoylation has been carried out on pentopyranosyl^{9,10} and hexopyranosylamines.² This paper describes our findings regarding the regioselective benzoylations of 2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose¹¹ (1), which is a D-glucosamine derivative suitable for N-deprotection.^{2,11}

RESULTS AND DISCUSSION

The results from the treatment of 2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (1) with 2-5.5 mol of benzoyl chloride in pyridine at different temperatures are shown in Table I. Although the work-up of these reactions involves a distribution between water and chloroform (see Experimental), TLC of the residue of the aqueous layer after concentration showed no presence of benzoyl derivatives 2-6. Compounds 2-4 each have a free anomeric hydroxyl group. However, they do not mutarotate (see Experimental), and 3 and 4 on acetylation yield exclusively the α -acetates 7 and 8, respectively. Additionally, the 1-O-benzoyl derivatives 5 and 6 have the α anomeric configuration (see below). These facts are in agreement with data reported for 2-(2-acylvinylamino)-2-deoxy- α -D-glucopyranose and their tetra-O-acetyl derivatives which indicate that the N-(2-acylvinyl) group reinforces the anomeric effect.¹¹ Moreover, it has also been described¹² that the catalytic de-O-acetylation at C-1 of 1,3,4,6-tetra-O-acetyl-2-(2-acylvinylamino)-2-deoxy- α - and β -D-glucopyranoses yielded only 3,4,6-tri-O-acetyl-2-(2-acylvinylamino)-2-deoxy- α -D-glucopyranoses.



	1	2	3	4	5	6	7	8
R ¹	H	H	H	H	Bz	Bz	Ac	Ac
R ²	H	H	Bz	H	Bz	Bz	Bz	Ac
R ³	H	H	H	Bz	H	Bz	Ac	Bz
R ⁴	H	Bz	Bz	Bz	Bz	Bz	Bz	Bz

The partially protected compounds which can be isolated with best yields are 2 (Table 1, entry 2), 3 and 5 (Table 1, entry 3). The experiments at low temperature (entries 1-3) indicate that the order of reactivity on benzylation of 1 is OH-6>OH-3>OH-1≈OH-4. The experiments at r.t. and 45 °C (entries 4 and 5) show that no significant regioselectivity takes place over 0 °C. The primary hydroxyl group is esterified more readily than are secondary hydroxyl groups, as is described for hexopyranosylamines² and hexopyranosides.¹³ The low reactivity of OH-4 has been observed in glucose,¹⁴ α-glucosides⁴ and glucosylamines,² and may be due to steric hindrance through a *gauche* interaction with CH₂OBz (considering that HO-6 is esterified first) and BzO-3. The relatively low reactivity of the anomeric hydroxyl group is attributed to its axial position^{2,9,11} and to the steric effect of the bulky enamino group on C-2. The HO-3 in 1 is less reactive than the same hydroxyl group in glucosylamines,² which may be explained by the above mentioned steric effect of the enamino group.

The acetyl derivatives 7 and 8 were prepared in order to complete the structural study of 3 and 4 respectively.

The structures of 2-8 were based on UV, IR, ¹H and ¹³C NMR, analytical and/or mass spectroscopic data.

Compounds 2-8 showed low stretching frequencies for NH (3230-3290 cm⁻¹) and C=O (1647-1674 cm⁻¹) groups, indicative of a chelated structure.^{11,15} This fact was also confirmed by the chemical shifts (see Tables 2-4) for the resonances of the NH (9.08-9.32 ppm) and one C=O group (167.9-169.8 ppm) and by the high value for $J_{\text{NH},\text{=CH}}$ (12.7-14.6 Hz, Table 3) which is indicative of antiperiplanar protons.

The values of $J_{1,2}$ (≈3.7 Hz), the high and positive optical rotation for 2-8 and the ¹H δ values for the 1-OAc signal (≈2.21 ppm) in 7-8 showed α(D) configuration for prepared compounds. The $^3J_{\text{H,H}}$ values for 3-8 indicated the

Table 1. Benzoylations of 1.

Entry	Temp. (°C)	[BzCl]/1 in mol	Benzoylated derivative and yield (%)						Benzoylated positions (%)		
			2	3	4	5	6	OH-6	OH-4	OH-3	OH-1
1	-40	2:1	31.0	1.2	0.5	---	---	32.7	0.5	1.2	---
2	-15	3:1	37.0	2.0	0.4	0.6	---	40.0	0.4	2.6	0.6
3	0	4:1	---	15.0	5.0	16.0	16.0	52.0	21.0	47.0	32.0
4	r.t.	9:2	---	---	---	3.0	80.0	83.0	80.0	83.0	83.0
5	45	11:2	---	---	---	---	92.0	92.0	92.0	92.0	92.0

Table 2. ¹H NMR chemical shifts (δ , ppm) of compounds **2-8** in CDCl₃.

Comp.	Sugar ring										Enamino group		
	H-1	H-2	H-3	H-4	H-5	H-6	H-6'	NH	=CH	CH ₂	CH ₃		
2a	5.13dd	<-----3.50-3.10m----->	5.64t	3.82t	4.30- 3.80m	4.53dd	4.40dd	9.08dd	8.00d	4.10q 4.04q	1.21t 1.19t		
3b	5.47d	3.61td	5.64t	3.82t	4.38m	4.73dd	4.63dd	9.14dd	8.02d	4.14q 4.04q	1.25t 1.17t		
4b	5.40bd	3.47td	4.20t	5.34t	4.53ddd	4.63dd	4.34dd	9.30dd	8.10d	4.19q 4.11q	1.30t 1.24t		
5b	6.53d	3.85td	5.69t	3.88m	4.20m	4.88dd	4.51dd	9.21dd	8.00d	4.08q 4.05q	1.18t 1.17t		
6b	6.61d	4.10- 3.90m	6.03t	5.79t	<-----4.65-4.39m----->	9.23dd	7.96d	7.96d	4.12q 4.10- 3.90m	1.18t 1.15t			
7b	6.24d	3.74td	5.59t	5.34t	4.21m	<-4.48-4.32m->	9.08dd	7.78d	4.15q 3.93q	1.12t 1.06t			
8b	6.24d	3.67td	5.53t	5.43t	4.35m	<-4.57-4.40m->	9.15dd	7.93d	4.15q 4.10q	1.20t 1.17t			

a. In (CD₃)₂SO.b. In CDCl₃.

Table 3. ^1H NMR coupling constants (Hz) of compounds 2-8.

Comp.	Sugar ring					Enamino group				
	$J_{1,2}$	$J_{2,3}$	$J_{3,4}$	$J_{4,5}$	$J_{5,6}$	$J_{5,6'}$	$J_{6,6'}$	$J_{\text{NH},=\text{CH}}$	$J_{2,\text{NH}}$	$^3J_{\text{H},\text{H}}$ (Et)
2a	3.7	---	---	---	2.5	5.0	12.5	14.3	9.0	7.0
3b	3.5	10.0	10.0	10.0	3.5	2.3	12.3	14.3	10.0	7.1
4b	3.7	9.7	9.7	9.7	2.7	4.3	11.9	13.7	9.7	7.1
5b	3.7	10.0	10.0	---	3.5	2.2	12.3	12.7	9.2	7.1
6b	3.8	9.9	9.9	9.9	---	---	---	14.6	9.6	7.1
7b	3.7	9.8	9.8	9.8	---	---	---	13.3	9.8	7.1
8b	3.7	9.7	9.7	9.7	---	---	---	13.1	9.7	7.1

a. In $(\text{CD}_3)_2\text{SO}$.b. In CDCl_3 .

Table 4. ^{13}C NMR chemical shifts (δ , ppm) for the sugar rings of compounds 2-8.

Comp.	C-1	C-2	C-3	C-4	C-5	C-6
2 ^a	90.7	63.4	71.6	69.7	70.2	64.2
3 ^b	91.5	62.5	74.8	69.4	70.3	63.2
4 ^b	91.4	63.9	71.6	70.6	67.7	62.9
5 ^b	91.6	61.3	73.8	68.5	73.0	62.8
6 ^b	91.4	61.4	71.2	68.4	70.0	62.1
7 ^b	90.6	61.1	71.2	67.6	70.0	61.8
8 ^b	90.3	60.7	70.6	68.5	69.9	62.2

a. In $(\text{CD}_3)_2\text{SO}$.

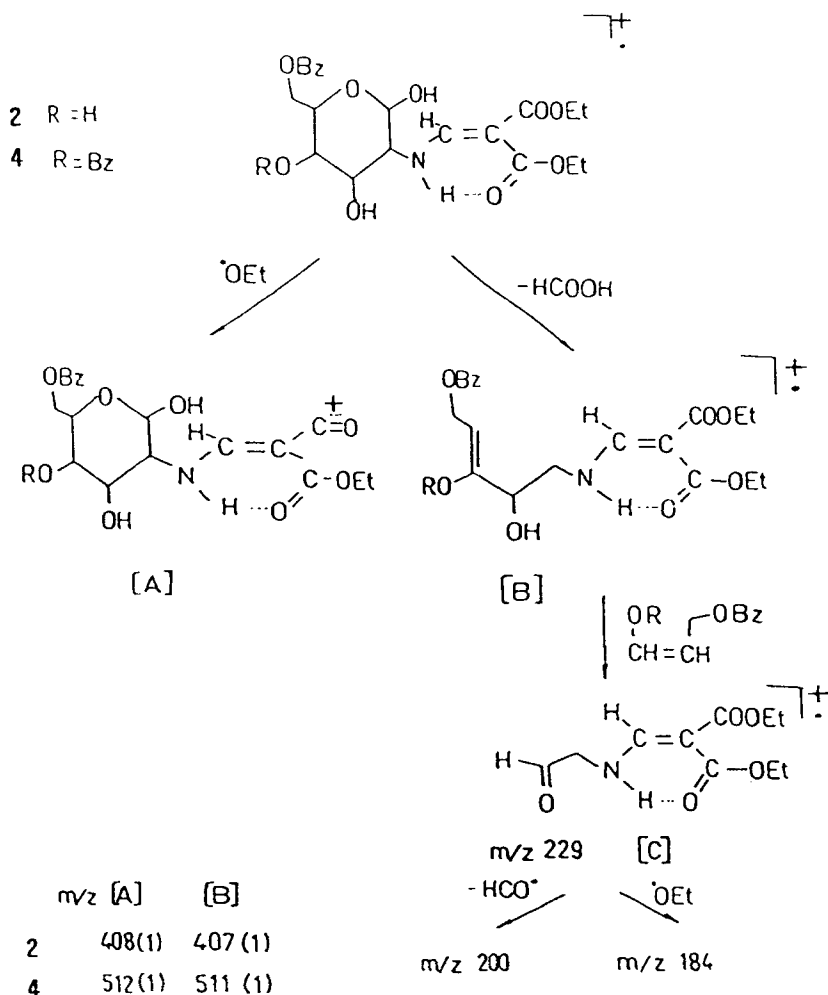
b. In CDCl_3 .

presence of a ${}^4\text{C}_1(\text{D})$ conformation in chloroform- d_3 solutions.

No transbenzoylations were observed when solutions of 2 (methyl sulphoxide- d_6), and 3-5 (chloroform- d_3) were kept for ten days at room temperature.

The mass spectra of 2-4, 7 and 8 showed molecular ions. The base peaks for 2-8 were always m/z 105 (Bz^+) and prominent fragments at m/z 122 (BzOH^+) and 77 (Ph^+) were observed. Scheme 1 summarises the primary fragments and the pathways of fragmentation for 2 and 4. The spectrum of 4 was carried out with a resolution of 10,000 (10% valley definition) and the atomic composition of the fragments agrees with the indicated structures. The loss of EtO^+ (peak A) is similar to that reported for

SCHEME 1



N-diethoxycarbonylvinyl glycosylamines.⁹ Peak B is evidence of the free anomeric hydroxyl group, and its structure is similar to that described for acetylated aldopyranosylamines.¹⁶ Peak C shows the presence of a hydroxyl group on C-3. Although no mass spectral data for acylated 2-deoxy-2-enamino aldoses are available, the results for 2-8 (see Experimental) agree with those reported for acetylated^{16,17} and benzoylated⁹ *N*-substituted glycosylamines.

EXPERIMENTAL

General procedures. Melting points are uncorrected. Optical rotations were measured at 20 ± 2 °C with a Perkin-Elmer 141 MC polarimeter, using 1 cm cell. UV spectra were recorded on a Beckman DU-7 spectrophotometer, IR spectra for KBr discs were measured with a Bomem Michelson MB-120 FT IR spectrophotometer. ^1H NMR (200.13 MHz) and ^{13}C NMR (50.33 MHz) were obtained with a Varian XL-200 spectrometer for solutions in CDCl_3 or DMSO-d_6 . Assignments of ^1H signals were confirmed by decoupling and H/D exchange experiments. Proton-decoupled APT¹⁸ (attached proton test) spectra were used to assist in the assignment of signals. EI mass spectra (70 eV) were recorded with a Kratos MS-80RFA instrument, with an ionising current of 100 μA , an accelerating voltage of 4 KV, and a resolution of 1000 (10% valley definition). The elemental compositions of the ions were determined with a resolution of 10000 (10% valley definition). Preparative chromatography was performed on Silica Gel 60 (Merck, 230 mesh), and TLC was carried out on silica gel 30 F₂₅₄ (Merck) plates.

Benzoylation of 2-Deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (1). (a) To a stirred solution of 1 (3.0 g, 8.6 mmol) in pyridine (15 mL) at -40 °C was gradually added benzoyl chloride (2.42 g, 2 mL, 17 mmol) in pyridine (5 mL). The mixture was kept for 72 h at -40 °C, then poured into ice-water (≈ 300 mL) to give a colourless syrup which was extracted with chloroform (3 \times 30 mL), and successively washed with 1M sulphuric acid (3 \times 30 mL), saturated aqueous sodium hydrogencarbonate (3 \times 30 mL) and water (3 \times 30 mL). The organic layer was dried (MgSO_4) and concentrated to dryness to give a syrup. Column chromatography (carbon tetrachloride-acetone 4:1 to 1:1) of this syrup gave 4,6-di-O-benzoyl-2-deoxy-2[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (**4**; 25 mg, 0.5%), R_F (carbon tetrachloride-acetone 2:1), 0.61; 3,6-di-O-benzoyl-2-deoxy-2[(2,2-diethoxycarbonylvinyl)amino]- α -D-

glucopyranose (3, 58 mg, 1.2%), $R_F=0.55$; and 6-O-benzoyl-2-deoxy-2[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (2, 1.224 g, 31%), $R_F=0.38$.

Compound 2 had mp 148-150 °C (from dichloromethane); $[\alpha]_D^{21} +82.2^\circ$ (c 1.0, dichloromethane), no mutarotation was observed after 24 h; UV (CH_2Cl_2) 232 and 280 nm (ϵ_{mM} 13.2 and 18.4); IR 3550 (OH), 3290 (NH), 1724 (C=O free), 1674 (C=O bonded), 1614 (C=C and NH),^{11,15} and 1271 cm^{-1} (C-O-C); ^1H NMR ($\text{DMSO}-d_6$) Tables 2 and 3, and δ 8.12-7.50 (m, 5H, Bz), 7.10 (d, 1H, $J_{1,\text{OH}}$ 5.0 Hz, OH-1), 5.41, and 5.40 (2d, 2H, $J_{3,\text{OH}}=J_{4,\text{OH}}=6.0$ Hz, OH-3, OH-4); ^{13}C NMR ($\text{DMSO}-d_6$) Table 4 and δ 168.4, 165.8, 165.3 (3C=O), 160.0 (=CH), 133.5-128.9 (6C, Ph), 88.3 (=C), 59.0, 58.9 (2 CH_2), 14.6 and 14.5 (2 CH_3). MS (scheme 1): m/z 453 (1, M^+), 408 (1, M^+-EtO^+ , [A]), 407 (2, M^+-HCOOH , [B]), 229 (17, [C]), 200 (40, [C]- HCO^+), 184 (19, [C]- EtO^+), 154 (73, 184- H_2CO), 122 (80, BzOH^+), 105 (100, Bz^+), and 77 (63, Ph^+).

Anal. Calcd for $\text{C}_{21}\text{H}_{27}\text{O}_{10}\text{N}$: C, 55.62; H, 6.00; N, 3.08. Found: C, 55.23; H, 5.96; N, 2.87.

Compound 3 was an amorphous and hygroscopic solid, and had $[\alpha]_D^{20} +120.3^\circ$ (c 0.8, dichloromethane), no mutarotation was observed after 24 h; UV (CH_2Cl_2) 239.5 and 279.0 nm (ϵ_{mM} 11.9 and 12.7); IR 3364 (OH), 3280 (NH), 1723 (C=O free), 1661 (C=O bonded), and 1271 cm^{-1} (C-O-C); ^1H NMR (CDCl_3) Tables 2 and 3, and δ 8.04-7.40 (m, 10H, 2Bz), 5.2 (bs, 2H, 2OH), ^{13}C NMR (CDCl_3) Table 4 and δ 168.8, 167.3, 166.8, 165.4 (4C=O), 158.9 (=CH), 133.5-128.3 (12C, 2Ph), 90.9 (=C), 60.2, 59.6 (2 CH_2), 14.2 and 14.0 (2 CH_3). MS: m/z 557 (1, M^+), 226 (6, Bz_2O^+), 122 (78, BzOH^+), 105 (100, Bz^+), and 77 (32, Ph^+); Found M^+ 557.1907. $\text{C}_{28}\text{H}_{31}\text{O}_{11}\text{N}$ requires M , 557.1867.

Compound 4 was an amorphous and hygroscopic solid, and had $[\alpha]_D^{21} +26.2^\circ$ (c 0.9, dichloromethane), no change in optical rotation was observed after 24 h; UV (CH_2Cl_2) 233 and 280 nm (ϵ_{mM} 15.4 and 11.3); IR 3356 (OH), 3260 (NH), 1724 (C=O free), 1661 (C=O bonded), 1603 (C=C and NH), and 1265 cm^{-1} (C-O-C); ^1H NMR (CDCl_3) Tables 2 and 3, and δ

8.12–7.25 (m, 10H, 2Bz), 4.51 (bs, 2H, 2OH); ^{13}C NMR (CDCl_3) Table 4 and δ 169.2, 166.1, 166.0, 165.9 (4C=O), 159.6 (=CH), 133.5–128.3 (12C, 2Ph), 90.4 (=C), 60.0, 59.7 (2CH_2), 14.3, and 14.1 (2CH_3). MS (scheme 1): m/z 557 (1, M^+), 512 (1, $\text{M}^+ - \text{EtO}^+$, [A]), 511 (1, $\text{M}^+ - \text{HCOOH}$, [B]), 229 (8, [C]), 200 (18, [C]- HCO^+), 184 (9, [C]- EtO^+), 154 (28, $184 - \text{H}_2\text{CO}$), 122 (38, BzOH^+), 105 (100, Bz^+), and 77 (30, Ph^+); Found M^+ 557.1900. $\text{C}_{28}\text{H}_{31}\text{O}_{11}\text{N}$ requires M , 557.1867.

(b) When the reaction was performed with 1 (0.5 g, 1.43 mmol) and benzoyl chloride (0.6 g, 0.5 mL, 4.30 mmol) in pyridine (5 mL) at -15°C as in (a), 2 (37%), 3 (2.0%), 4 (0.4%), and 1,3,6-tri-*O*-benzoyl-2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (5, 6 mg, 0.6%, R_f 0.71), were obtained.

Compound 5 had mp $194\text{--}195^\circ\text{C}$ (from ethanol), $[\alpha]_D^{19} + 146.5^\circ$ (c 0.4, dichloromethane); UV (CH_2Cl_2) 236.0 and 287.5 nm (ϵ_{mM} 27.1 and 16.6); IR 3435 (OH), 3230 (NH), 1736, and 1723 (C=O free), 1657 (C=O bonded), 1603 (C=C and NH), and 1269 cm^{-1} (C-O-C); ^1H NMR (CDCl_3) Tables 2 and 3, and 8.18–7.39 (m, 15H, 3Bz), 3.63 (bs, 1H, OH-4); ^{13}C NMR (CDCl_3) Table 4 and δ 168.9, 166.5, 165.8, 165.2, 164.3 (5C=O), 158.4 (=CH), 134.2–128.4 (18C, 3Ph), 92.6 (=C), 60.1, 59.8 (2CH_2), 14.3 and 14.1 (2CH_3). MS: m/z 253(3), 208(1), 207(1), 180 (8), 122 (52, BzOH^+), 105 (100, Bz^+), and 77 (66, Ph^+).

Anal. Calcd for $\text{C}_{35}\text{H}_{35}\text{O}_{12}\text{N}$: C, 63.53; H, 5.33; N, 2.11. Found: C, 63.24; H, 5.66; N, 2.06.

(c) When the reaction was carried out from 1 (0.5 g, 1.43 mmol) and benzoyl chloride (0.64 g, 0.53 mL, 4.58 mmol) at 0°C as in (a), 3 (15%), 4 (5%), 5 (16%), and 1,3,4,6-tetra-*O*-benzoyl-2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (6, 0.143 g, 16%, R_f 0.79) were obtained.

Compound 6 had mp $181\text{--}182^\circ\text{C}$ (from ethanol), $[\alpha]_D^{22} + 219.2^\circ$ (c 1.0, dichloromethane); UV (CH_2Cl_2) 239.5 and 276 nm (ϵ_{mM} 30.4 and 21.0); IR 3260 (NH), 1740 (C=O free), 1661 (C=O bonded), 1609 (C=C and NH) and 1260 cm^{-1} (C-O-C);

^1H NMR (CDCl_3) Tables 2 and 3, and δ 8.27–7.27 (m, 2OH, 4Bz); ^{13}C NMR (CDCl_3) Table 4 and δ 167.9, 165.8, 165.6, 165.0, 164.8, 164.2 (6C=O), 158.0 (=CH), 134.1–128.2 (24C, 4Ph), 92.8 (=C), 59.9, 59.5 (2CH₂), 14.1 and 14.0 (2CH₃). MS: m/z 244(2), 122 (85, BzOH⁺), 105 (100, Bz⁺), and 77 (32, Ph⁺).

Anal. Calcd for C₄₂H₃₉O₁₃N: C, 65.88; H, 5.13; N, 1.83. Found: C, 65.93; H, 5.20; N, 1.89.

(d) To a stirred solution of 1 (2 g, 5.73 mmol), in pyridine (8 mL) at 0 °C was added gradually benzoyl chloride (3.62 g, 3.0 mL, 25.76 mmol). After 48 h at room temperature, the mixture was poured into ice-water (\approx 300 mL), and the crude product was treated as in (a) to give 5 (3.0%) and 6 (80%).

In each reaction (a)–(d) the absence of benzoyl derivatives 2–6 in the aqueous layer was confirmed by TLC of the residue of this layer after concentration to dryness.

(e) To a solution of 1 (0.5 g, 1.43 mmol) in pyridine (5 mL) was added benzoyl chloride (1.10 g, 0.91 mL, 7.86 mmol). After 24 h at 45 °C, the mixture was poured into ice-water (\approx 100 mL), and the crude product was crystallised from ethanol to give 6 (1.0 g, 92%).

1,4-Di-O-acetyl-3,6-di-O-benzoyl-2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (7) and **1,3-Di-O-acetyl-4,6-di-O-benzoyl-2-deoxy-2-[(2,2-diethoxycarbonylvinyl)amino]- α -D-glucopyranose (8)**. Conventional treatments¹⁹ of 3 and 4 (0.070 g, 0.125 mmol) with pyridine (3 mL) and acetic anhydride (0.025 g, 0.023 mL, 0.251 mmol) gave 7 (0.025 g, 31%) and 8 (0.073 g, 90%) respectively as pale and hygroscopic syrups. Compound 7 had $[\alpha]_{\text{D}}^{20} +154.8^\circ$ (c 0.7, dichloromethane); UV 233 and 278 nm (ϵ_{mM} 18.2 and 14.7); IR 3250 (NH), 1755, 1723 (C=O free), 1655 (C=O bonded), 1603 (C=C and NH), 1273 and 1237 cm⁻¹ (C–O–C); ^1H NMR (CDCl_3) Tables 2 and 3, and δ 7.99–7.29 (m, 10H, 2Bz), 2.21 (s, 3H, 1-OAc) and 1.87 (s, 3H, AcO-4); ^{13}C NMR (CDCl_3) Table 4 and δ 169.2, 165.8, 165.4, 164.9, 164.3, 163.8 (6C=O), 158.1 (=CH), 133.5–128.2 (12C, 2Ph), 93.0

(=C), 60.0, 59.6 (2CH₂), 20.7, 20.4 (2CH₃CO), 14.3 and 14.1 (2CH₃). MS: *m/z* 641 (2, M⁺), 581 (2, M⁺-AcOH), 536 (1, M⁺-Bz⁺), 508 (1, 581-CO₂Et), 459 (3, 581-BzOH), 430 (2), 249 (3), 270 (5), 122 (32, BzOH⁺), 105 (100, Bz⁺), 77 (27, Ph⁺), 60 (12, AcOH⁺), and 43 (30, Ac⁺); Found M⁺ 641.2080. C₃₂H₃₅O₁₃N requires M, 641.2108. Compound 8 had [α]_D²² +54.7° (c 1.0, dichloromethane); UV 233 and 278 nm (ε_{mM} 17.6 and 15.8); IR 3260 (NH), 1753, 1726 (C=O free), 1660 (C=O bonded), 1609 (C=C and NH), 1256 and 1239 cm⁻¹ (C-O-C); ¹H NMR (CDCl₃) Table 2 and 3, and δ 8.00-7.34 (m, 10H, 2Ph), 2.22 (s, 3H, AcO-1) and 1.85 (s, 3H, 3-OAc); ¹³C NMR (CDCl₃) Table 4 and δ 169.8, 168.6, 168.4, 165.9, 165.3, 165.0 (6C=O), 158.6 (=CH), 133.6-128.2 (12C, 2Ph), 93.0 (=C), 60.7, 60.1 (2CH₂), 20.7, 20.3 (2CH₃CO), 14.26 and 14.10 (2CH₃); MS: *m/z* 641 (2, M⁺), 596 (1, M⁺-EtO⁺), 581 (2, M⁺-AcOH), 508 (1, 581-CO₂Et), 399 (1, 581-BzOH-AcOH), 249 (2), 180 (5), 122 (45, BzOH⁺), 105 (100, Bz⁺), 77 (37, Ph⁺), and 43 (18, Ac⁺); Found M⁺ 641.2098. C₃₂H₃₅O₁₃N requires M, 641.2108.

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